Molecular biogeography of the fungus-dwelling saproxylic beetle *Bolitophagus reticulatus* indicates rapid expansion from glacial refugia

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Running title: Biogeography of *Bolitophagus reticulatus*

ABSTRACT

The geographic distributions of species associated with European temperate broadleaf forests were significantly influenced by glacial-interglacial cycles. These species persisted the glacial periods in Mediterranean and extra-Mediterranean refugia and expanded northwards during the interglacial stages. The common saproxylic beetle *Bolitophagus reticulatus* closely depends on European temperate broadleaf forests. The beetle mostly develops in the tinder fungus *Fomes fomentarius*, a major decomposer of broadleaf-wood. We sampled *B. reticulatus* in sporocarps from European (*Fagus sylvatica*) and Oriental beech (*F. orientalis*) across Europe and the Caucasus region. We analysed mitochondrial gene sequences (*cox1, cox2, cob*) and microsatellites to reconstruct the geographic distribution of glacial refugia and postglacial recolonization pathways. We found only marginal genetic differentiation of *B. reticulatus*, except for a significant split between populations of the Caucasus region and Europe. This indicates the existence of past refugia south of the Great Caucasus, and a contact zone with European populations at the Crimea region. Further, potential refugia might have been located at the foothills of the Pyrenees and the Balkan region. Our genetic data suggest a phalanx-wise recolonization of Europe, which reflects the high mobility of this beetle species.

Keywords: Broadleaf forest, *Fomes fomentarius*, biogeography, genetic analysis, refugia, expansion, phalanx-wise, mobility

INTRODUCTION

The glacial-interglacial cycles of the Pleistocene caused severe range shifts of most species across Europe (Hewitt, 1999; 2000; Schmitt, 2007; Schmitt & Varga, 2012). Many European species persisted the past glacial periods in Mediterranean refugia (Hewitt, 1999), as well as in extra-Mediterranean refugia of Central Europe (Schmitt & Varga, 2012). Also the ponto-caspian area was proposed as potential glacial refugium for European taxa (Tarkhnishvili *et al.*, 2012, Neiber & Hausdorf, 2015). These range modifications resulted in inter- and intraspecific genetic signatures, such as differentiation through long-term isolation in disjunct glacial refugia (Hewitt, 2000). Also range expansions after glacial periods are reflected in the genetics of species. They follow two propagation patterns: a pioneer process (with the two types, stepping stone wise and leptokurtic), implying repeated founder effects in the wake of population expansions into new habitat patches (Ibrahim *et al*., 1996). This propagation pattern creates typical signatures of gradual loss of genetic diversity in the course of colonization (Ibrahim et al. 1996). In contrast, phalanx-wise colonization implies area-wide expansion, and therefore a lack of genetic signatures along colonization routes (Hewitt, 2000).

The biogeography of broadleaf tree species has been intensively studied during the past years (Pott, 2000; Brunet *et al*., 2010). Forests dominated by broadleaves currently occur in diverse ecoregions and include the Atlantic, Central European, Balkan, Baltic, Dinaric, and Caucasus mixed forests, which are equipped with typical plant, fungus, and animal species (Brunet *et al*., 2010; Müller *et al*., 2013). They have persisted in disjunct glacial refugia. Tree species with high cold tolerances, such as birch (*Betula* sp.), occurred in extra-Mediterranean and northern refugia during the glacial stages (Svenning, *et al.*, 2008; Giesecke *et al.*, 2017). More thermophilic tree species, such as European beech (*Fagus sylvatica*), persisted the glacial stages in various disjunct Mediterranean refugia, as well as in a number of cryptic extra-Mediterranean refugia along the edge of the Eastern Alps, the Balkan Peninsula and northern

Spain (Magri *et al*., 2006, 2008; Saltré *et al.,* 2013). After the last glacial period, the European beech recolonized central and northern Europe mainly from the Balkan region (Magri *et al*., 2006), while the populations in the western Mediterranean area, such as northern Spain, played a rather minor role as potential sources for recolonization (Magri *et al*., 2006, 2008; Saltré *et al.*, 2013).

While the biogeographic history of all tree species forming the European broadleaf forests is well studied (Magri *et al*., 2006, 2008; Svenning, *et al.*, 2008; Saltré *et al.,* 2013; Giesecke *et al.*, 2017), comparably little data and evidence on the biogeographic history of animal species relying on European broadleaf forests are available (Stauffer *et al*., 1999; Rukke, 2000; Pons *et al.*, 2011; Drag *et al*., 2011, 2015, 2018; Jiménez-Alfaro *et al*., 2018). Moreover, in many of these studies the Caucasus region is not considered, although further refugia could have been located in this region.

In this study we analysed the genetic structure of the darkling beetle *Bolitophagus reticulatus* (Linnaeus, 1767) (Tenebrionidae, Tenebrionini, Bolitophagini), a typical representative of the fauna of European broadleaf forest. The larvae and adults live in polypores and mostly inhabit the tinder fungus *Fomes fomentarius* (L.) Fr. 1849 (Midtgaard *et al.*, 1998, 2013; Nilsson, 1997). The beetle species is widespread across the Palaearctic region and very mobile (Jonsson, 2003). We sampled individuals of this species across its Western Palaearctic distribution range, including the Caucasus region. We analysed mitochondrial DNA sequences and polymorphic microsatellites allowing the investigation at different rates of evolution. Based on these data we identify past glacial refugia and range expansions during interglacial periods. In particular we aim to answer the following questions:

1. Do the refugial areas of *B. reticulatus* correspond to the refugia of tree species being part of the European broadleaf forest?

2. What is the role of the Caucasian region in the context of glacial survival and postglacial recolonization of the Western Palaearctic?
3. How did post-glacial range expansion take place, pioneer- or phalanx-wise?
4. Do genetic structures coincide with the ecology and behaviour of *B. reticulatus*?

MATERIAL AND METHODS

Study species

The genus *Bolitophagus* is represented in the Palearctic by a total of four species (Iwan *et al.*, 2020). The most widespread species is *Bolitophagus reticulatus*, having a Palearctic distribution, but being absent from the central Mediterranean. Its larvae and adults live in polypores and are among the most frequent inhabitants of the tinder fungus *Fomes fomentarius* (Friess *et al.*, 2019). Adults of the beetle feed on spores from living basidiocarps, but are also commonly found in dead and deteriorated polypores, where its larvae develop (Midtgaard *et al.*, 1998, 2013). Experimental studies found single individuals to fly up to 125 km in a flight mill experiment (Jonsson, 2003). This high mobility is also supported by local scale studies (Jonsson *et al*., 2003; Zytynska *et al*., 2018). The main host of *B. reticulatus* is *F. fomentarius* (Nilsson, 1997), but it was also recorded from other polypores (e.g. *Phellinus nigricans*, *Fomitopsis pinicola*, *Piptoporus betulinus*, *Ganoderma applanatum*, *Laetiporus sulphureus* and *Daedaleopsis* spp.; Bouget *et al.*, 2019). *F. fomentarius* occurs on a range of broadleaf tree species, mostly beech (*Fagus* spp.) and birch (*Betula* spp.), but rarely also others like oak (*Quercus* spp.) and maple (*Acer* spp.).

Sampling

We collected 281 individuals of *B. reticulatus* from 57 beech forest sites across major parts of the beetle´s western Palaearctic distribution range, including the Caucasus region. We sampled five individuals at each site (wherever possible). Sampling was conducted during the years 2014, 2015, and 2017. We extracted the individuals from sporocarps of *F. fomentarius* and subsequently stored them in 99% ethanol until further analyses. An overview of all sampling sites including GPS coordinates is compiled in Appendix Table S1. All individuals used in this study are stored at the Terrestrial Ecology Research Group, Technical University Munich (TUM), Freising, Germany.

151

152 **Molecular analyses**

153 DNA was extracted from head, thorax and fore legs applying the Qiagen DNeasy kit (Qiagen,
154 Hilden, Germany) based on the standard protocol for tissue samples. Partial mitochondrial
155 genes cytochrome oxidase subunit I (*cox1*), cytochrome c oxidase subunit II (*cox2*), and
156 cytochrome b (*cob*) were amplified using the primer combinations and PCR conditions
157 described in Rangel López *et al*. (2018). Successfully amplified PCR products were purified
158 with ExoSap (Thermo Fischer Scientific) and subsequently sequenced in both directions by
159 the Genomics Service Unit (GSU) of the Ludwig-Maximilians-Universität München (LMU),
160 Germany. We successfully generated *cox1*, *cox2*, and *cob* sequences for 208 individuals (out
161 of the 281 individuals sampled). An overview of all sequences and GenBank accession
162 numbers are given in Appendix Table S2.

163

164 We successfully genotyped seventeen polymorphic microsatellites for 255 individuals (out of
165 the 281 individuals sampled) (Appendix Table S2), with the same primers and conditions
166 successfully applied in a previous study (Zytynska *et al*., 2018). We used two multiplex
167 combinations, each with 8-9 primer pairs, using three fluorescent dyes: 6-FAM, HEX, and
168 TAmRA, alongside the ROX size standard. PCR products were run on an ABI 3130xl Genetic
169 Analyzer (Applied Biosystems – Life Technologies GmbH, Darmstadt, Germany) at the GSU
170 of the LMU, Germany. Further details on protocols applied are given in Zytynska et al.
171 (2018).

172

173 **Phylogenetic and demographic analyses**

174 Forward and reverse reads of mtDNA sequences were assembled with GENEIOUS v. 6.1.8
175 (https://www.geneious.com). After removing primer sequences and low-quality base calls

176 from the sequence ends, multiple sequence alignment was performed per marker using the
177 MUSCLE (Edgar, 2004 a, b) algorithm as implemented in GENEIOUS.
178
179 Mitochondrial haplotypes were extracted from the aligned mitochondrial supermatrix in
180 PEGAS v. 0.13 (Paradis, 2010). Individuals with more than 100 missing sites were excluded
181 and sites with missing or ambiguous data were disregarded. Haplotype networks were inferred
182 using an infinite sites model (i.e. uncorrected distance) with PEGAS and the spatial distribution
183 of haplotypes was mapped with a combination of the *R*-packages MAPS v. 3.3.0 (Becker *et al*.,
184 2018), RASTER v. 3.1-5 (Hijmans, 2020), and GGPLOT2 v. 3.3.0 (Wickham, 2016).
185
186 A phylogenetic tree was inferred with IQ-TREE v. 2.0-rc2 (Minh *et al*., 2020). *Nalassus*
187 *laevioctostriatus*, *Opatrum sabulosum*, and *Eledonoprius armatus* were chosen as outgroup
188 based on an already published phylogeny of tenebrionid beetles (Kergoat et al., 2014).
189 Respective sequences were obtained from NCBI GenBank (Appendix Table S2). Data was
190 partitioned into the three genes (*cox1*, *cox2*, *cob*) and their codon positions for a total of 9
191 initial partitions used as input for MODELFINDER (Kalyaanamoorthy *et al*., 2017). This
192 approach not only selects the best fitting substitution model for each partition, but also merges
193 initial partitions according to their statistical properties to reduce parameter space. The top ten
194 percent of partition pairs were evaluated (option *-rcluster 10*). The heuristic tree search was
195 repeated 10 times. The best tree was chosen and rooted with *Opatrum sabulosum*. 1×10^5
196 ultrafast bootstrap replicates were performed to provide branch support (Hoang *et al*., 2018).
197
198 We performed Coalescent Bayesian Skyline analysis (Drummond *et al.*, 2005) with BEAST
199 v. 2.6.2 (Boukaert et al., 2014). Outgroups were excluded for this analysis. An estimate of the
200 *cox1* substitution rate in tenebrionid beetles (3.54 ± 0.38 % My^{-1}) (Papadopoulou *et al.*, 2010)
201 was used to calibrate the mitochondrial tree in time, using the mean estimate with a relaxed

lognormal molecular clock model. Optimal models of nucleotide substitution and partition scheme were inferred with MODELFINDER (Kalyaanamoorthy *et al*., 2017) in IQ-TREE; initial partitions were set to the three genes. The topology was linked across genes. Three independent MCMCs were run for 8×10^7 generations, with sampling every 5×10^3 generations. Convergence of independent runs to similar values, stationarity, and effective sample sizes were assessed in TRACER v. 1.7.1 (Rambaut *et al*., 2018) after removing a burn-in of 25 % of samples. Based on the combined post-burn-in sample of all three runs, Bayesian Skyline plots were generated with TRACER and *ggplot2* v. 3.3.0 (Wickham, 2016). The posterior sample of trees was summarized with TREEANNOTATOR from the BEAST software package, using maximum clade credibility and common ancestor heights.

Analyses of population structure

Analyses of population structure were done with microsatellite data using R v. 4.0.2 (R Core Team, 2019) in R-STUDIO v. 1.2.1335 (RStudio Team, 2018). Mean F_{ST}, G_{ST}, G'_{ST}, and D_{Jost} were calculated as basic descriptive molecular statistics of population differentiation per locus. Allelic richness and number of unique allele combinations, as well as mean observed and mean expected heterozygosity were calculated using the packages POPPR v. 2.8.5 (Kamvar *et al*., 2014; 2015), DIVERSITY v. 1.9.90 (Keenan *et al*., 2013), and ADEGENET v. 2.1.2 (Jombart, 2008; Jombart & Ahmed, 2011). Pairwise F_{ST}-values were calculated for clusters inferred from total evidence (see below) using ADEGENET.

Populations of *B. reticulatus* were inferred with GENELAND v. 4.9.2 (Guillot et al. 2005b, 2012). GENELAND applies mixture models to infer clusters that are in Hardy-Weinberg equilibrium with linkage equilibrium between loci. We inferred genetic clusters using the uncorrelated frequency model based on three datasets: (1) microsatellites, (2) mitochondrial sequences, and (3) the combination thereof (referred to as total evidence in the following).

SNPs were extracted from mitochondrial sequences using ADEGENET. The algorithm considers geographic coordinates of samples, assuming that populations are spatially separated and experience little gene flow (spatial model) (Guillot et al. 2005a). A spatial jitter of 0.00001 degree was applied to avoid fixation of samples from one locality in the same cluster. MCMC chains were run for one million generations (five million for mtDNA), sampling every 1000^{th} generation (5000^{th} for mtDNA). Each analysis was repeated three times to ensure stability of results. Log likelihood and log posterior density trace plots were inspected to ensure convergence and stationarity of runs and to identify potential outliers that were stuck in local optima using CODA v. 0.19-3. The maximum number of populations was set to 50, which roughly corresponds to sampling localities. The maximum rate of the Poisson process was set to the number of individuals in the respective dataset. The maximum number of nuclei in the Poisson-Voronoi tessellation was set to two times the number of individuals, which is suggested for analyses under the spatial model. Null alleles were not filtered. Posterior samples of each repeat run were separately summarized using the *PostProcessChain*-function after removing a burn-in of 100,000 generations (400,000 for the total evidence dataset).

To test for isolation-by-distance, geographic distances were transformed from geographical coordinates to meters using the RASTER package (Hijmans 2020). Genetic distances were calculated using ADEGENET (Jombart, 2008; Jombart & Ahmed, 2011). The distances were plotted against each other for all pairs of sampling locations and complemented by a two-dimensional density extrapolation to explore potential geographically and genetically isolated populations. Correlation of the distance matrices was statistically tested by a Mantel test as implemented in ADE4 (v. 1.7-15; Chessel *et al*., 2004). Significance was assessed by 10,000 randomizations.

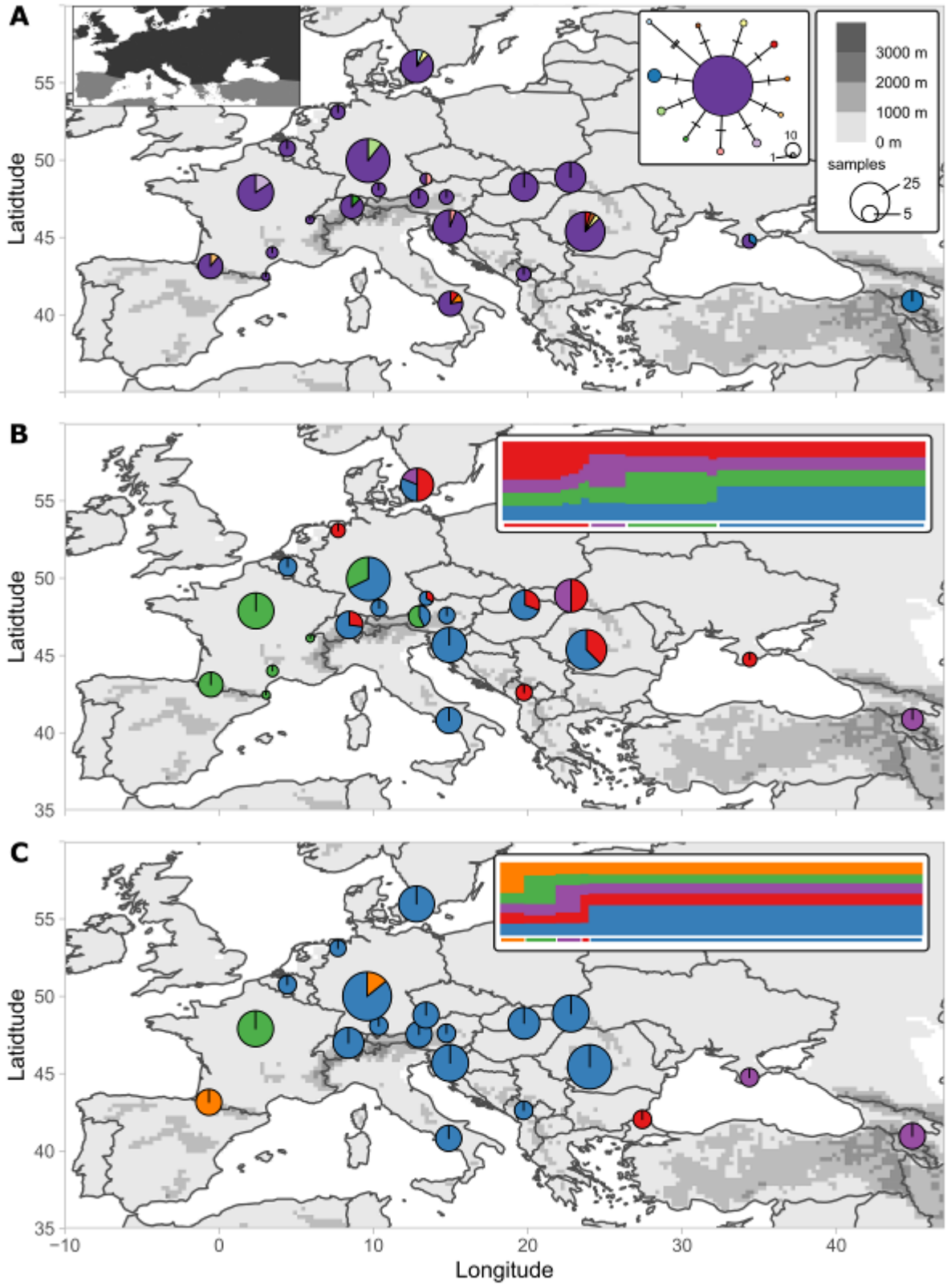

Figure 1: Spatial distribution of (A) 12 mitochondrial haplotypes and haplotype network (*cox1, cox2,* and *cob*), (B) four mitochondrial genetic clusters from GENELAND analysis, and (C) five nuclear genetic clusters from GENELAND analyses of

polymorphic microsatellites. The size of the pie charts represents the number of samples; the size of the circles in the haplotype network represents the number of respective haplotypes. Pie charts may summarize several close by localities. The inset in (A) shows the approximate distribution of *Bolitophagus reticulatus* in the study area (own data and www.gbif.org). Insets in (B) and (C) show membership probabilities (y-axis) of individuals (x-axis) to inferred clusters, which are colour coded for the respective maps. Coloured bars below the plot indicate assigned group membership.

RESULTS

The total concatenated alignment of the three mitochondrial genes consisted of 208 individuals and 1,605 bp (*cox1*: 525 bp, *cox2*: 626 bp, *cob*: 454 bp). Missing data was 1.26%, 2.98%, and 1.19% for *cox1*, *cox2*, and *cob*, respectively. Variation in the mitochondrial genes was generally low (Appendix Fig. S1). The combined mitochondrial genes differed at twelve segregating sites (excluding sites with missing or ambiguous data), resulting in twelve mitochondrial haplotypes (haplotype diversity = 0.24, nucleotide diversity = 0.00024; 167 haplotypes were found using all sites with pairwise deletion of missing and ambiguous data). One haplotype was noticeably dominant in terms of individual number (in 87% of all individuals) and distribution range. This haplotype represented the centre of a star-shaped haplotype network (Fig. 1A, dark violet haplotype). Other, less frequent haplotypes were regionally restricted with two exceptions that both occurred in the Carpathian Basin (Fig. 1A, yellow and red haplotypes). GENELAND identified four mitochondrial clusters which considerably overlapped geographically (Fig. 1B). The Crimean population represented a combination of haplotypes of the Caucasian and European haplotypes. This pattern was also discovered with Bayesian and Maximum Likelihood phylogenetic analyses.

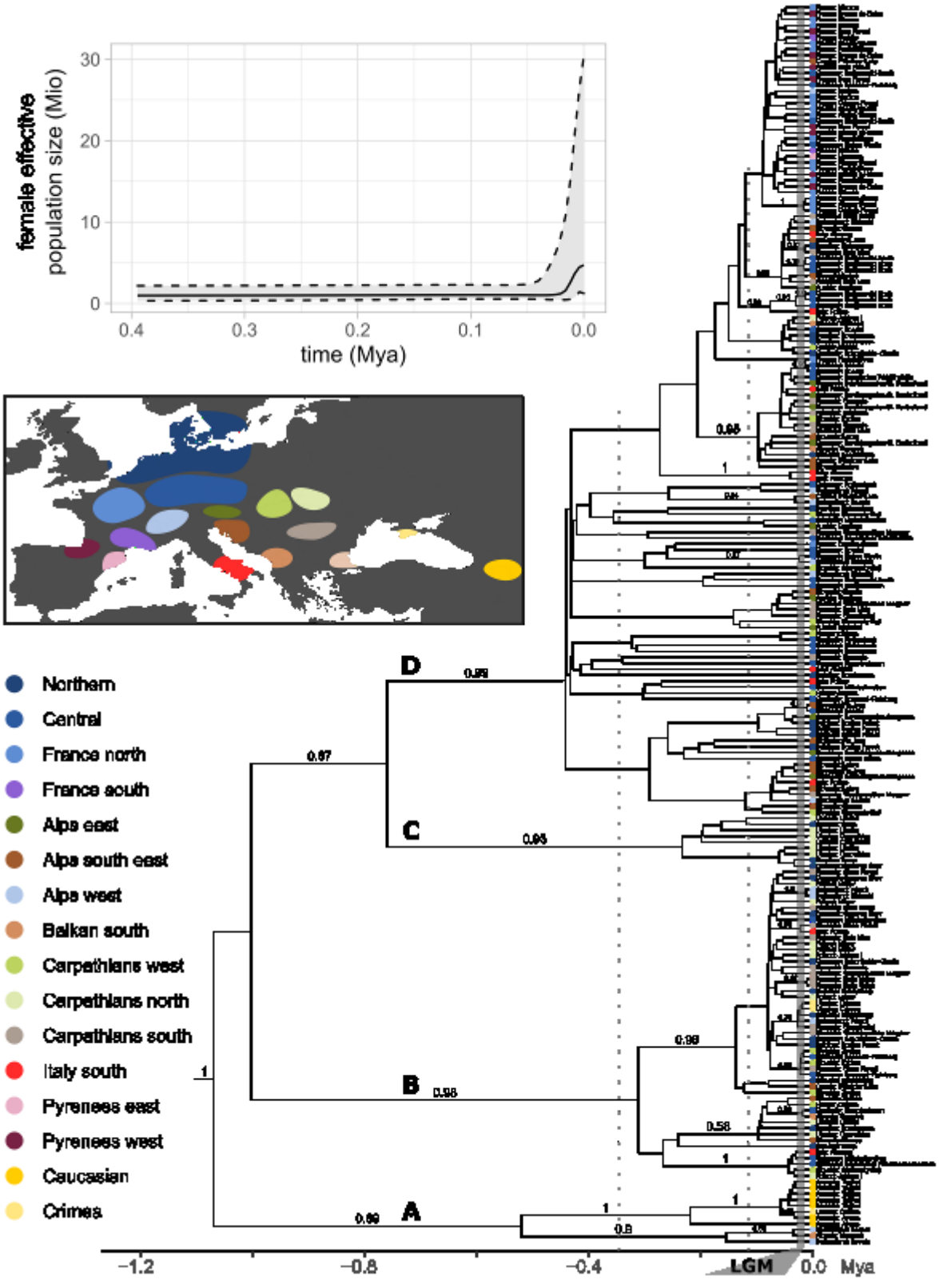

Figure 2: Time calibrated phylogenetic tree from Bayesian analysis of mitochondrial DNA sequences (*cox1, cox2,* and *cob*). Vertical dotted lines indicate the onset of the last glacial periods. The vertical grey line marks the last glacial maximum. Colour dots

286 at the trees' tips illustrate geographic origin of the sample. Coloured areas on the map
287 roughly encircle sampling points of the present study and include refugia of European
288 and Oriental beech. Upper Inset: Bayesian Skyline plot showing demographic change
289 in female effective population size over time, assuming a generation time of one year.
290
291 The dated mitochondrial tree from Bayesian inference suggested four major and mostly well
292 supported clades (Fig. 2 clades A–D). An accumulation of recent diversification events was
293 detected around 100 kya. Geographic turnover was high, so that specimens from the same
294 region were rarely restricted to a single clade. The exception were the Caucasus specimens
295 from Armenia, the easternmost sampling locality, which clustered in one clade together with
296 one specimen from Crimea. Clades A and B largely included populations from more eastern
297 locations and showed a connection to the most eastern records of *B. reticulatus* (Ukraine,
298 Armenia). Those clades also comprised specimens from eastern and northern Europe (Fig. 2).
299 Clade C was largely restricted to the northern Carpathians, but likewise included specimens
300 from Denmark and Sweden. Clade D comprised most specimens, originating from all over
301 Europe except the far eastern localities on Crimea and Armenia. Nearly all populations
302 sampled across France assembled into one lineage (part of clade D), only interspersed with
303 three specimens from neighbouring German sites and one from Plitvice Lakes (Croatia).
304 Posterior supports and crown diversification ages are given in Table 2. The mitochondrial tree
305 from the maximum likelihood search largely confirmed the genetic clades obtained from
306 Bayesian analysis, although some topological differences with little support were present
307 (Appendix Fig. S1, clades A–D). Bayesian Skyline analysis showed a marked increase in
308 population size since 20 kya, with a recent tendency to reduced growth approximately 5 kya
309 (Fig. 2).
310

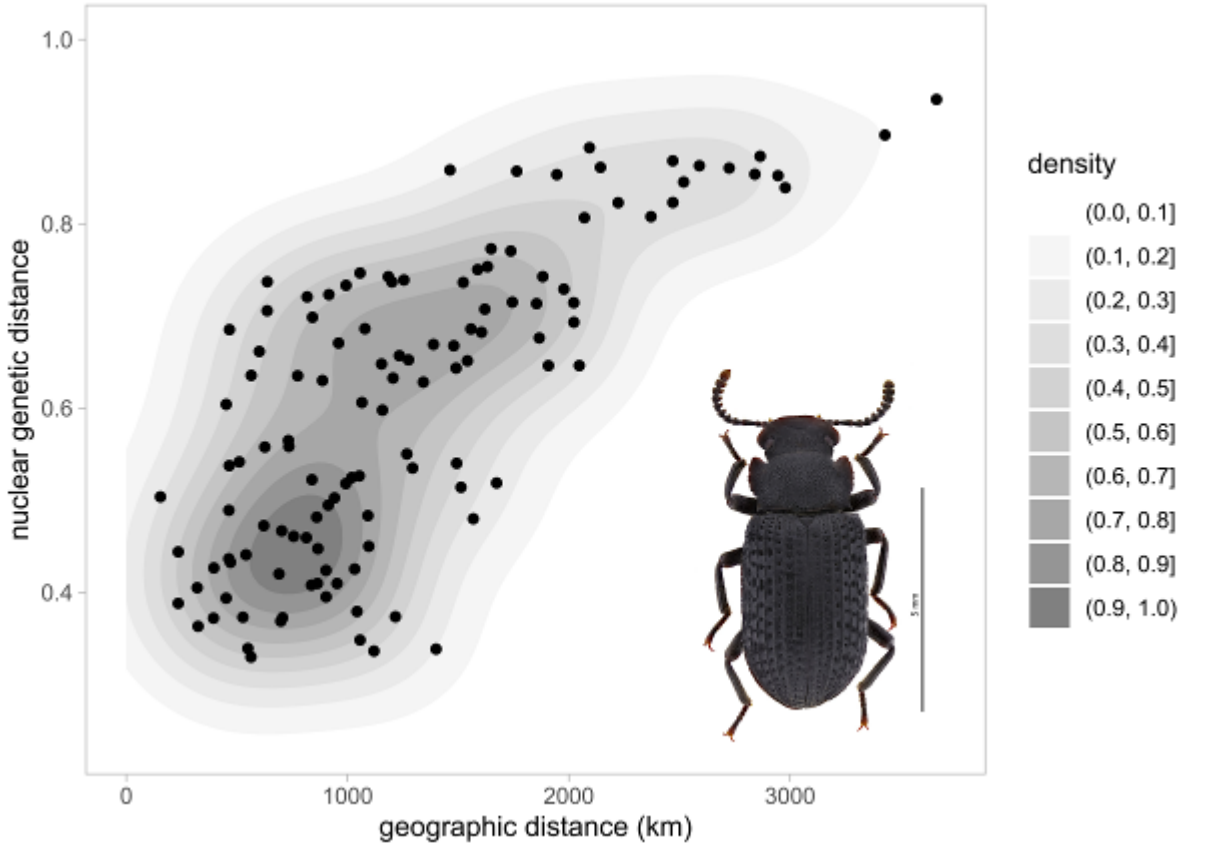

Figure 3: Pairwise geographic and genetic distance between localities illustrate isolation by distance. Nuclear genetic distance was inferred from microsatellite data. Shades of grey indicate data point density.

Similar to our results obtained from mitochondrial data, global statistics of microsatellite data revealed generally low genetic diversity (Appendix Table S3). Observed and expected heterozygosity were 0.46 and 0.78, respectively, on average across loci. Mean F_{ST} was 0.15, mean G_{ST} 0.28, mean G'_{ST} 0.64, and mean D_{Jost} 0.50. One nuclear cluster dominated the genetic structure based on polymorphic microsatellites (Fig. 1C; blue cluster). However, individual cluster composition differed substantially between mitochondrial and nuclear inferences. In contrast to results from mitochondrial sequences, nuclear clusters were spatially well separated. One exception was a disjunct Pyrenean-German cluster. This is in line with low population differentiation indices that were found between three clusters inferred from

total evidence (mitochondrial, nuclear, and geography): the F_{ST} value between a western and an eastern population was 0.14, while genetic exchange between them and a large central European cluster seemed to be substantial, resulting in F_{ST} values < 0.05 (Appendix Figure S2). Furthermore, we found significant correlation of genetic and geographic distances among localities based on the Mantel test (expectation from simulation: -0.002, variance: 0.039, observation: 0.738, $p < 0.001$).

DISCUSSION

The study of three mitochondrial genes and polymorphic microsatellites allowed us to reconstruct the postglacial dispersal pathways of *B. reticulatus*. Except for the European-Caucasian split, which may be due to common isolation of beetle and host tree, we found very little genetic differentiation. This is most likely explained by genetic depletion in glacial refugia and rapid postglacial dispersal out of these refugia.

The European-Caucasian split

The clade restricted to the Caucasus region was clearly distinguishable from the European clade based on mitochondrial DNA and microsatellite analyses. These genetic signatures suggest the existence of a refuge area south of the Great Caucasus. This finding goes in line with previous molecular biogeographic studies on other species where European and Caucasian populations were included (see e.g., Filipova-Marinova, 1995; Pavlova *et al.*, 2005; Hansson *et al*., 2008). Molecular analyses identified a sister species relationship of European beech (*F. sylvatica*) and Oriental beech (*F. orientalis*) (Renner *et al*., 2016), which are distributed in Europe and the Caucasus, respectively (www.euforgen.org; accessed December 2020). The same isolating forces that caused the intraspecific differentiation in *B. reticulatus* might have caused speciation in the two beech species. Furthermore, our data indicated the Crimean region being the contact zone between European populations and the populations of the Great Caucasus. Previous studies also identified the Crimean region as important contact zone, e.g. for land snails (Neiber & Hausdorf, 2017).

Refugia across Central Europe

Infrequent mitochondrial haplotypes occurred regionally restricted, with two exceptions, both for the Carpathian Basin. This suggests glacial refugia of *B. reticulatus* on the Balkan Peninsula, and postglacial range expansions across the south-eastern European region, with

major areas on the Balkan Peninsula, including the foothills of the Carpathians and areas of central Europe. This scenario was also supported by phylogenetic inference, and goes in line with a range of previous studies (reviewed in Schmitt 2007). The postglacial range expansions from the Balkan Peninsula across major parts of eastern central Europe coincides with the phylogeography of the European meadow grasshopper *Chorthippus parallelus* (Lunt, Ibrahim, & Hewitt, 1998), which is name giving to one of the three paradigms stated by Hewitt (Hewitt 1996, 1999, 2000).

Given the generally high spatial admixture that was evident from mitochondrial DNA in *B. reticulatus*, it is noteworthy that phylogenetic analyses recovered all but two specimens from France in one clade, although not well supported (Fig. 2). Likewise, GENELAND clustered all individuals from France into one cluster when used with mtDNA data and suggested a connection to central Europe (Fig. 1B). Potential scenarios shaping such pattern are extra-Mediterranean glacial refugia located at the Massif Central or at the foothills of the Pyrenees with subsequent postglacial range expansion. This is a frequently observed pattern in biogeographic studies of organisms of temperate Europe (e.g., Schmitt & Seitz, 2001). However, the low genetic diversity in France indicates a bottleneck effect during the last glacial maximum and thus likely small refugia. The multiple extra-Mediterranean glacial refugia found for *B. reticulatus* (e.g. the Carpathian Basin, Massif Central/Pyrenees) go in line with findings for various European broadleaf tree species which are used by *F. fomentarius*. For example, molecular data of the European beech also indicate past refugia at the foothills of the Pyrenees (Magri 2008). Other studies underline that various tree species of the European broadleaf forest expanded early after the last glacial maximum northwards, or even survived in more northern extra-Mediterranean refugia (Chlebicki & Lorenc, 1997; Svenning *et al.*, 2008; Schmitt & Varga, 2012).

384 **Range expansions**

385 The observed genetic structures supported that *B. reticulatus* occurred restrictively in areas
386 with beech-dominated broadleaf forest providing good conditions for the tinder fungus
387 (Schwarze, 1994). The tinder fungus and *B. reticulatus* are capable dispersers and likely
388 exhibit similar post-glacial population expansions. The chronogram in our study indicated an
389 accumulation of diversification events in *B. reticulatus* with the beginning of the last ice-age
390 around 100 kya; the median estimate of the onset of population growth was ca. 20 kya, which
391 coincides with the end of the last glacial maximum. This population growth pattern of *B.*
392 *reticulatus* inferred from mtDNA suggests range expansions and population increase before
393 the period of colonization by beech trees, derived from paleontological evidence (Magri *et al*.,
394 2008). This is an indication that the colonization of the tinder fungus and *B. reticulatus* might
395 have occurred independently of the beech, and took place much earlier. A plausible scenario
396 is the expansion together with birch trees, although today in temperate Europe beech is the
397 main host for the polypore.
398
399 The weak genetic differentiation, alongside high geographic turnover of mtDNA and low F_{ST}
400 values among regional clusters, as well as the lack of gradual loss of genetic diversity along
401 potential colonization pathways, allows the inference of the expansion pattern of *B.*
402 *reticulatus*. While pioneer processes lead to signatures of gradual loss of genetic diversity in
403 the course of colonization (Ibrahim et al. 1996), phalanx-wise colonization results in a lack of
404 genetic signatures along colonization routes (Hewitt, 2000). The observed lack of genetic
405 differentiation, in combination with the isolation by distance pattern, approves a phalanx-wise
406 colonization of Europe and reflects the strong mobility of *B. reticulatus* (Jonsson, 2003).
407 Similar genetic signatures were also found for the longhorn beetle *Rosalia alpina* (Drag *et al.*,
408 2018), which inhabits a similar habitat. Our data underline the capability of *B. reticulatus* to

409 rapidly colonize new habitats and the frequent individual exchanges among local populations,
410 which counteracts potential genetic differentiation.

Data Availability Statement

The mtDNA sequences underlying this article are available in the GenBank Nucleotide Database at www.ncbi.nlm.nih.gov/genbank/, and can be accessed with the accession numbers MH383529–MH383770 for *cob*, MH383771–MH384020 for *cox1*, and MH384021–MH384258 for *cox2*. Sequence alignments and phylogenetic trees are available in TreeBase at http://purl.org/phylo/treebase/phylows/study/TB2:S27736. Microsatellite data are available in the online supplementary material (Appendix Table S2).

Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

Table S1: Sampling sites and genetic diversity measures.

Table S2: GenBank Accesion numbers for mtDNA sequences and microsatellite data.

Table S3: Global statistics of microsatellite data.

Figure S1. Mitochondrial tree from maximum likelihood analysis.

Figure S2. Spatial distribution of three clusters inferred by GENELAND based on total evidence.

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References

Becker RA, Wilks AR (Original S code), Brownrigg R, Minka TP, Deckmyn A (R version). 2018. maps: Draw Geographical Maps. R package version 3.3.0. https://CRAN.R-project.org/package=maps

Bouget C, Brustel H, Noblecourt T, Zagatti P. 2019. *Les Coléoptères saproxyliques de France – Catalogue écologique illustré*, Muséum national d'Histoire naturelle, Paris, 744p. (Patrimoines naturels; 79).

Boukaert R, Heled J, Kühnert D, Vaughan T, Wu CH, Xie D, Suchard MA, Rambaut A & Drummond AJ. **2014**. BEAST 2: A Software Platform for Bayesian Evolutionary Analysis. *PLOS computational biology* **10**: 1–6.

Brunet J, Fritz Ö & Richnau G. 2010. Biodiversity in European beech forests – a review with recommendations for sustainable forest management. *Ecological Bulletin* **55**: 77–94.

Chessel D, Dufour A & Thioulouse J. **2004**. The ade4 Package – I: One-Table Methods. *R News* **4**: 5–10.

Chlebicki A & Lorenc MW. **1997**. Subfossil Fomes fomentarius from a Holocene fluvial deposit in Poland. *The Holocene* **7**: 101–103.

Drag L, Hauck D, Bérces S, Michalcewicz J, Jelaska LS, Aurenhammer S, Cizek L. 2015. Genetic differentiation of populations of the threatened saproxylic beetle Rosalia Longicorn, *Rosalia alpina* (Coleoptera: Cerambycidae) in Central and South-east Europe. *Biological Journal of the Linnean Society* **116:** 911–925.

Drag L, Hauck D, Pokluda P, Zimmermann K, Cizek L. 2011. Demography and dispersal ability of a threatened saproxylic beetle: A mark-recapture study of the Rosalia longicorn (*Rosalia alpina*). *PLoS One* **6:** e21345.

Drag L, Hauck D, Rican O, Schmitt T, Shovkoon DF, Godunko RJ, Curletti G, Cizek L. 2018. Phylogeography of the endangered saproxylic beetle Rosalia longicorn *Rosalia alpina* (Coleoptera, Cerambycidae), corresponds with its main host, the European beech (*Fagus sylvatica*, Fagaceae). *Journal of Biogeography* **45:** 2631–2644.

Drummond AJ, Rambaut A, Shapiro B & Pybus OG. **2005**. Bayesian Coalescent Inference of Past Population Dynamics from Molecular Sequences. *Molecular Biology and Evolution* **22**: 1185–1192.

Edgar RC. **2004a**. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* **32**: 1792–1797.

Edgar RC. **2004b**. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* **5**: 113.

Filipova-Marinova M. 1995. Late Quaternary history of the Genus *Fagus* in Bulgaria. In Bozilova E & Tonkov S (eds). *Advances in Holocene Palaeoecology in Bulgaria*. Pensoft Publishers.

Friess N, Müller JC, Abrego N, Aramendi P, Bässler C, Bouget C, Brin A, Bussler H, Georgiev K, Gil R, Gossner MM, Heilmann-Clausen J, Isaacson G, Krištín A, Lachat T, Larrieu L, Los S, Magnanou E, Maringer A, Mergner U, Mikolas M, Opgenoorth L, Schmidl J, Svoboda M, Thorn S, Vrezec A, Vanderkhoven K, Winter B, Wagner T, Zapponi L, Brandl R & Seibold S. 2019. The species-rich arthropod communities in fungal fruitbodies are weakly structured by climate and biogeography across European beech forests. *Diversity and Distributions* **25**: 783-796.

Giesecke T, Brewer S, Finsinger W, Leydet M & Bradshaw RHW. **2017**. Patterns and dynamics of European vegetation change over the last 15,000 years. *Journal of Biogeography* **44**: 1441–1456.

Guillot G, Estoup A, Mortier F, Cosson JF. 2005a. A spatial statistical model for landscape genetics. *Genetics* **170**: 1261–1280.

Guillot G, Mortier F, Estoup A. 2005b. Geneland: A program for landscape genetics. *Molecular Ecology Notes* **5**: 712–715**.**

Guillot G, Renaud S, Ledevin R, Michaux J, Claude J. 2012. A Unifying Model for the Analysis of Phenotypic, Genetic and Geographic Data. *Systematic Biology*, 61: 897–911.

Habel JC, Mulwa RK, Gassert F, Rödder D, Ulrich W, Borghesio L, Husemann M & Lens L. **2014**. Population signatures of large-scale, long-term disjunction and small-scale, short-term habitat fragmentation in an Afromontane forest bird. *Heredity* **113**: 205–214.

Hansson B, Hasselquist D, Tarka M, Zehtindjiev P & Bensch S. **2008**. Postglacial Colonisation Patterns and the Role of Isolation and Expansion in Driving Diversification in a Passerine Bird. *PLOS ONE* **3**: e2794.

Hewitt GM. 1996. Some genetic consequences of ice ages, and their role in divergence and speciation. *Biological Journal of the Linnean Society* **58**, 247–276.

Hewitt GM. 1999. Post-glacial recolonization of European biota. *Biological Journal of the Linnean Society* **68:** 87–112.

Hewitt GM. 2000. The genetic legacy of the Quaternary ice ages. *Nature* **405:** 907–913.

Hijmans RJ. 2020. raster: Geographic Data Analysis and Modelling. R package version 3.1-5. https://CRAN.R-project.org/package=raster

Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. 2018. UFBoot2: Improving the Ultrafast Bootstrap Approximation. *Molecular Biology and Evolution* **35:** 518–522.

Ibrahim KM, Nichols RA, Hewitt GM. 1996. Spatial patterns of genetic variation generated by different forms of dispersal during range expansion. *Heredity* **77**: 282–291.

Iwan D, Löbl I, Bouchard P, Bousquet Y, Kamiński M, Merkl O, Ando K & Schawaller W. 2020. Family Tenebrionidae Latreille, 1802. In: Iwan D, Löbl I, eds. *Catalogue of Palaearctic Coleoptera, Volume 5, Tenebrionoidea*. Revised and updated second edition. Leiden: Brill NV, 969 pp.

Jiménez-Alfaro B, Girardello M, Chytrý M, Svenning JC, Willner W, Gégout JC, Agrillo E, Campos JA, Jandt U, Kącki Z, Šilc U, Slezák M, Tichý L, Tsiripidis I, Turtureanu PD, Ujházyová M & Wohlgemuth T. 2018. History and environment shape species pools and community diversity in European beech forests. *Nature Ecology & Evolution* **2**: 483–490.

Jombart T. 2008. adegenet: a R package for the multivariate analysis of genetic markers. *Bioinformatics* **24:** 1403-1405.

Jombart T, Ahmed I. 2011. adegenet 1.3-1: new tools for the analysis of genome-wide SNP data. *Bioinformatics*.

Jonsson M. 2003. Colonisation ability of the threatened tenebrionid beetle *Oplocephala haemorrhoidalis* and its common relative *Bolitophagus reticulatus*. *Ecological Entomology* **28:** 159–167.

Jonsell M, Schroeder M, Larsson T. 2003. The saproxylic beetle *Bolitophagus reticulatus*: its frequency in managed forests, attraction to volatiles and flight period. *Ecography* **26**: 421 – 428.

Jonsson M, Johannesen J & Seitz A. **2003**. Comparative genetic structure of the threatened tenebrionid beetle *Oplocephala haemorrhoidalis* and its common relative *Bolitophagus reticulatus*. *Journal of Insect Conservation* **7**: 111–124.

Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS. 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nature Methods* **14:** 587–589.

Kamvar ZN, Tabima JF, Grünwald NJ. 2014. Poppr: an R package for genetic analysis of populations with clonal, partially clonal, and/or sexual reproduction. *PeerJ 2:* e281.

546 **Kamvar ZN, Brooks JC and Grünwald NJ. 2015.** Novel R tools for analysis of genome-
547 wide population genetic data with emphasis on clonality. *Frontiers in Genetics* **6:** 208.
548 **Keenan K, McGinnity P, Cross TF, Crozier WW & Prodöhl PA. 2013.** diveRsity: An R
549 package for the estimation of population genetics parameters and their associated
550 errors. *Methods in Ecology and Evolution* **4**: 782–788.
551 **Kergoat GJ, Soldati L, Clamens AL, Jourdan H, Jabbour-Zahab R, Genson G,**
552 **Bouchard P & Condamine FL**. **2014**. Higher level molecular phylogeny of darkling
553 beetles (Coleoptera: Tenebrionidae). *Systematic Entomology* **39**: 486–499.
554 **Knutsen H, Rukke BA, Jorde PE & Ims RA**. **2000**. Genetic differentiation among
555 populations of the beetle *Bolitophagus reticulatus* (Coleoptera: Tenebrionidae) in a
556 fragmented and a continuous landscape. *Heredity* **84**: 667–676.
557 **Lunt DH, Ibrahim KM & Hewitt GM**. **1998**. mtDNA phylogeography and postglacial
558 patterns of subdivision in the meadow grasshopper *Chorthippus parallelus*. *Heredity*
559 **80**: 633–641.
560 **Magri D, Vendramin GG, Comps B, Dupanloup I, Geburek T, Gomory D, Latalowa M,**
561 **Litt T, Paule L, Roure JM, Tantau I, van der Knaap WO, Petit RJ & de Beaulieu**
562 **JL**. **2006**. A new scenario for the Quaternary history of European beech populations:
563 palaeobotanical evidence and genetic consequences. *New Phytologist* **171**: 199–221.
564 **Magri D**. **2008**. Patterns of post-glacial spread and the extent of glacial refugia of European
565 beech (*Fagus sylvatica*). *Journal of Biogeography* **35**: 450–463.
566 **Midtgaard F, Rukke BA & Sverdrup-Thygeson A**. **1998**. Habitat use of the fungivorous
567 beetle *Bolitophagus reticulatus* (Coleoptera: Tenebrionidae): Effects of basidiocarp
568 size, humidity and competitors. *European Journal of Entomology* **95**: 559–570.
569 **Minh BQ, Schmidt HA, Chernomor O, Schrempf D, Woodhams MD, von Haeseler A,**
570 **Lanfear R. 2020.** IQ-TREE 2: New models and efficient methods for phylogenetic
571 inference in the genomic era. *Molecular Biology and Evolution*
572 **Müller J, Brunet J, Brin A, Bouget C, Brustel H, Bussler H, Förster B, Isacsson G,**
573 **Köhler F, Lachat T & Gossner MM**. **2013**. Implications from large-scale spatial
574 diversity patterns of saproxylic beetles for the conservation of European Beech forests.
575 *Insect Conservation and Diversity* **6**: 162–169.
576 **Neiber MT & Hausdorf B**. **2015**. Phylogeography of the land snail genus *Circassina*
577 (Gastropoda: Hygromiidae) implies multiple Pleistocene refugia in the western
578 Caucasus region. *Molecular Phylogenetics and Evolution* **93**: 129–142.

Neiber MT & Hausdorf B. **2017**. Molecular phylogeny and biogeography of the land snail genus *Monacha* (Gastropoda, Hygromiidae). *Zoologica Scripta* **46**: 308–321.

Nilsson T. 1997. Survival and habitat preferences of adult *Bolitophagus reticulatus.* *Ecological Entomology* **22:** 82-89.

Papadopoulou A, Anastasiou I, Vogler AP 2010. Revisiting the insect mitochondrial molecular clock: The mid-aegean trench calibration. *Molecular Biology and Evolution* **27:** 1659–1672.

Paradis E. 2010. pegas: an R package for population genetics with an integrated-modular approach. *Bioinformatics* 26: 419-420.

Pavlova A, Zink RM, Rohwer S, Koblik EA, Red'kin YA, Fadeev IV & Nesterov EV. **2005**. Mitochondrial DNA and plumage evolution in the white wagtail *Motacilla alba*. *Journal of Avian Biology* **36**: 322–336.

Pons JM, Olioso G, Cruaud C & Fuchs J. **2011**. Phylogeography of the Eurasian green woodpecker (*Picus viridis*). *Journal of Biogeography* **38**: 311–325.

Pott R. 2000. Die Entwicklung der europäischen Buchenwälder in der Nacheiszeit. *Rundgespräche der Kommission für Ökologie*, **18**, 49-75.

R Core Team. 2019. R: A language and environment for statistical computing. R Foundation for statistical Computing, Vienna, Austria. URL https://www.R-project.org/.

Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. 2018. Posterior Summarization in Bayesian Phylogenetics Using Tracer 1.7. *Systematic Biology* **67:** 901–904.

Rangel López JÁ, Husemann M, Schmitt T, Kramp K & Habel JC. **2018**. Mountain barriers and trans-Saharan connections shape the genetic structure of *Pimelia* darkling beetles (Coleoptera: Tenebrionidae). *Biological Journal of the Linnean Society* **124**: 547–556.

Renner SS, Grimm GW, Kapli P, Denk T. 2016. Species relationships and divergence times in beeches: new insights from the inclusion of 53 young and old fossils in a birth–death clock model. *Philosophical Transactions of the Royal Society B:* **371:** 20150135.

RStudio Team. 2018. RStudio: Integrated Development for R. RStudio, Inc., Boston, MA URL http://www.rstudio.com/.

Rukke BA. 2000. Effects of habitat fragmentation: increased isolation and reduced habitat size reduces the incidence of dead wood fungi beetles in a fragmented forest landscape. *Ecography* **23:** 492–502.

612 **Saltré F, Saint-Amant R, Gritti ES, Brewer S, Gaucherel C, Davis BAS & Chuine I**.
613 **2013**. Climate or migration: what limited European beech post-glacial colonization?
614 *Global Ecology and Biogeography* **22**: 1217–1227.
615 **Schmitt T**. **2007**. Molecular biogeography of Europe: Pleistocene cycles and postglacial
616 trends. *Frontiers in Zoology* **4**: 11.
617 **Schmitt T, Varga Z. 2012.** Extra-Mediterranean refugia: The rule and not the exception?
618 *Frontiers in Zoology* **9:** 22.
619 **Schwarze F. 1994.** Wood rotting fungi: *Fomes fomentarius* (L.: Fr.) Fr.: Hoof or tinder
620 fungus. *Mycologist 8:* 32–34.
621 **Stauffer C, Lakatos F & Hewitt GM. 1999.** Phylogeography and postglacial colonization
622 routes of *Ips typographus* L. (Coleoptera, Scolytidae). *Molecular Ecology* **8**: 763–773.
623 **Svenning JC, Normand S & Kageyama M**. **2008**. Glacial refugia of temperate trees in
624 Europe: insights from species distribution modelling. *Journal of Ecology* **96**: 1117–
625 1127.
626 **Tarkhnishvili D, Gavashelishvili A & Mumladze L**. **2012**. Palaeoclimatic models help to
627 understand current distribution of Caucasian forest species. *Biological Journal of the*
628 *Linnean Society* **105**: 231–248.
629 **Wickham H. 2016.** ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York.
630 **Zytynska SE, Doerfler I, Gossner MM, Sturm S, Weisser WW, Müller J. 2018.** Minimal
631 effects on genetic structuring of a fungus-dwelling saproxylic beetle after
632 recolonisation of a restored forest. *Journal of Applied Ecology* **55:** 2933– 2943.
633

634 **Figures**

635 Figure 1: Spatial distribution of (A) 12 mitochondrial haplotypes and haplotype network
636 (*cox1, cox2,* and *cob*), (B) four mitochondrial genetic clusters from GENELAND analysis, and
637 (C) five nuclear genetic clusters from GENELAND analyses of polymorphic microsatellites.
638 The size of the pie charts represents the number of samples; the size of the circles in the
639 haplotype network represents the number of respective haplotypes. Pie charts may summarize
640 several close by localities. The inset in (A) shows the approximate distribution of
641 *Bolitophagus reticulatus* in the study area (own data and www.gbif.org). Insets in (B) and (C)
642 show membership probabilities (y-axis) of individuals (x-axis) to inferred clusters, which are
643 colour coded for the respective maps. Coloured bars below the plot indicate assigned group
644 membership.

645

646 Figure 2: Time calibrated phylogenetic tree from Bayesian analysis of mitochondrial DNA
647 sequences (*cox1, cox2,* and *cob*). Vertical dotted lines indicate the onset of the last glacial
648 periods. The vertical grey line marks the last glacial maximum. Colour dots at the trees' tips
649 illustrate geographic origin of the sample. Coloured areas on the map roughly encircle
650 sampling points of the present study and include refugia of European and Oriental beech.
651 Upper Inset: Bayesian Skyline plot showing demographic change in female effective
652 population size over time, assuming a generation time of one year.

653

654 Figure 3: Pairwise geographic and genetic distance between localities illustrate isolation by
655 distance. Nuclear genetic distance was inferred from microsatellite data. Shades of grey
656 indicate data point density.

657

658

659 **Tables**

660 Table 1: Investigated regions and genetic diversities obtained for all populations analysed.
661 Given are the coordinates in decimal format (WGS84), the number of mitochondrial DNA
662 samples n_{mt}, haplotypes *HT*, nuclear DNA samples n_n, alleles *A*, and allele combinations, as
663 well as observed and expected heterozygosity, H_o, and H_e respectively.

664

region	n_{mt}	*HT*	n_n	*A*	*Comb.*	H_o	H_e
Alps east	8	1	15	95	106	0.35	0.61
Alps south-east	17	2	20	146	195	0.50	0.77
Alps west	9	2	14	138	150	0.48	0.75
Balkan east	na	na	5	47	50	0.48	0.48
Balkan south	3	1	5	86	76	0.63	0.70
Carpathians north	14	1	20	129	175	0.48	0.70
Carpathians south	23	4	29	156	223	0.46	0.72
Carpathians west	12	1	15	97	122	0.44	0.65
Caucasian	7	1	10	38	43	0.16	0.24
Central	33	3	50	177	299	0.51	0.74
Crimea	3	2	5	49	51	0.39	0.48
France north	19	2	19	93	142	0.50	0.62
France south	2	1	na	na	na	na	na
Italy south	9	3	10	102	117	0.51	0.72
Northern	23	3	28	149	228	0.45	0.75
Pyrenees east	1	1	na	na	na	na	na
Pyrenees west	9	2	10	71	82	0.42	0.53

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666

667 Table 2: Crown ages of major lineages from Bayesian divergence dating (compare Fig. 2).

Node	Posterior clade support	Median age (kya)	Mean age (kya)	95 % HPD interval (kya)
Root	1.00	971.8	1070.5	398.8 – 1960.6
A	0.89	400.3	456.3	107.1 – 928.7
B	0.98	279.2	308.2	102.3 – 582.2
C + D	0.67	585.7	644.8	234.5 – 1181.8
C	0.93	174.5	216.1	24.2 – 515.5
D	0.99	389.2	434.0	153.8 – 818.5

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